


Metformin induced autophagy in diabetes mellitus – Tuberculosis co-infection patients: A case study

by Bernadette Dian Novita

FILE	2-METFORMIN_INDUCED_.PDF (456.77K)		
TIME SUBMITTED	15-FEB-2019 03:52PM (UTC+0700)	WORD COUNT	5861
SUBMISSION ID	1078613741	CHARACTER COUNT	29360

AUTHOR QUERY FORM

 ELSEVIER	Journal: IJTB Article Number: 262	Please e-mail your responses and any corrections to: E-mail: corrections.esch@elsevier.thomsondigital.com
---	--	--

Dear Author,

Please check your proof carefully and mark all corrections at the appropriate place in the proof (e.g., by using on-screen annotation in the PDF file) or compile them in a separate list. Note: if you opt to annotate the file with software other than Adobe Reader then please also highlight the appropriate place in the PDF file. To ensure fast publication of your paper please return your corrections within 48 hours.

For correction or revision of any artwork, please consult <http://www.elsevier.com/artworkinstructions>.

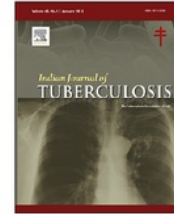
Any queries or remarks that have arisen during the processing of your manuscript are listed below and highlighted by flags in the proof. Click on the 'Q' link to go to the location in the proof.

Location in article	Query / Remark: click on the Q link to go Please insert your reply or correction at the corresponding line in the proof
Q1	The author names have been tagged as given names and surnames (surnames are highlighted in teal color). Please confirm if they have been identified correctly.
Q2	Highlights are in incorrect format. Hence the same has been deleted. It should consist of 3–5 bullet points (85 characters per bullet point, including spaces). Please edit the highlights to meet the requirement.
Q3	Please check abstract for correctness.
Q4	“Your article is registered as a regular item and is being processed for inclusion in a regular issue of the journal. If this is NOT correct and your article belongs to a Special Issue/Collection please contact s.bodatte@elsevier.com immediately prior to returning your corrections.”.
	<div data-bbox="396 1247 1016 1350"> Please check this box or indicate your approval if you have no corrections to make to the PDF file <input checked="" type="checkbox"/> </div>

Thank you for your assistance.

Available online at www.sciencedirect.com

ScienceDirect

journal homepage: <http://www.journals.elsevier.com/indian-journal-of-tuberculosis/>

Original article

Metformin induced autophagy in diabetes mellitus – Tuberculosis co-infection patients: A case study

Q1 Bernadette Dian Novita^{a,*}, Mulyohadi Ali^b, Agung Pranoto^c, Endang Isbandiati Soediono^a, Ni Made Mertaniasih^d

^a Department of Pharmacology and Therapy, Faculty of Medicine Widya Mandala Catholic University Surabaya, Indonesia

^b Department of Pharmacology, Faculty of Medicine, Brawijaya University, Indonesia

^c Department of Internal Medicine, Faculty of Medicine Airlangga University/Dr. Soetomo Hospital, Indonesia

^d Department of Clinical Microbiology, Faculty of Medicine Airlangga University/Dr. Soetomo Hospital, Indonesia

ARTICLE INFO

Article history:

Received 16 August 2017

Accepted 9 April 2018

Available online xxx

Keywords:

Type 2 diabetes mellitus-tuberculosis co-infection patients

Metformin

AFB smear reversion

Autophagy

ABSTRACT

Metformin (MET) is a potential combination drug to elevate anti-TB efficacy. However, the clinical effect, especially smear reversion, during metformin applied with anti-tuberculosis and insulin in patients with type 2 DM newly TB co-infection were remain unknown.

An observational clinical study was done in DM newly TB co-infection outpatients at Surabaya Paru Hospital. This study evaluated MET therapy, at least 2 months, accompanying with insulin and anti-TB regimens and compared to comparison group. The smear, microtubule associated Protein1 Light Chain 3B (MAP1LC3B) level, as the presentation of autophagy, Superoxide Dismutase (SOD) level, Interferon (IFN)- γ and Interleukin (IL)-10 levels were evaluated twice.

From 42 participants in this study, 22 participants of observation group that received additional MET therapy, 100% had sputum smear reversion after 2-months intensive phase of anti-TB therapy. Whereas 25% of 20 participants of comparison group did not undergo reversion inserts sputum smear.

As conclusion, MET has the potential of being an additive combination therapy to enhance the bactericidal effect of anti-TB on DM-TB coinfection patients. Metformin enhances the effects of anti-TB and insulin therapy in increasing the smear reversion by increasing autophagy.

© 2018 Tuberculosis Association of India. Published by Elsevier B.V. All rights reserved.

1. Introduction

Tuberculosis (TB) nowadays is one of “global health emergency” diseases.¹ The increasing evidence in TB

patients associated with the rising number of patients at risk for TB, i.e. patients with immunocompromised condition such patients with HIV, diabetes mellitus (DM), cancer and autoimmune diseases. The TB infection at risk in DM patients increased 2.39 times with the risk of failure of

* Corresponding author.

E-mail address: novita@ukwms.ac.id (B.D. Novita).

<https://doi.org/10.1016/j.ijtb.2018.04.003>

0019-5707/© 2018 Tuberculosis Association of India. Published by Elsevier B.V. All rights reserved.

Please cite this article in press as: Novita BD, et al. Metformin induced autophagy in diabetes mellitus – Tuberculosis co-infection patients: A case study, *Indian J Tuberc.* (2018), <https://doi.org/10.1016/j.ijtb.2018.04.003>

anti-TB therapy increased 1.69 times with the smear reversion $\pm 60\%$.^{2,3}

First-line anti-TB regimens, rifampicin, isoniazid (INH), ethambutol, pyrazinamide, use as anti-TB, thus, those etiology therapy aimed for curing the patients, preventing death, preventing recurrence, cutting off the transmission chains and preventing pathogen resistance by eradication *Mycobacterium tuberculosis*.^{4–6} The effectiveness of anti-TB are also influenced by the host immune response due to the interaction of anti-TB. Adjunctive therapy with immunomodulators that enhance TB immunity (host-directed therapy, HDT) could shorten treatment durations and improve TB outcomes.^{7–9} The identification of new host-directed therapies aimed at improving the clinical outcome of TB patients is a priority for TB management and WHO, including some drugs that work on immune regulation. Currently, several studies have been conducted to determine the function of some immunomodulatory agents, including corticosteroids, TNF blockers, thalidomide, and non-steroidal anti-inflammatory (NSAIDs) as adjunct of OAT therapy.^{8,9}

One of HDT mechanism is autophagy due to its ability in inhibiting the TB infection process.^{8,9} The process of activating autophagy from formation to maturation and then fusion with lysosomes for phagolysosome or autophagy processes requires many activators and protein (ATGs), one of the proteins representing phagolysosome or autophagy is MAP1LC3B/ATG8.

Metformin hydrochloride (MET), biguanide the oral anti diabetic agent, recently, by a comprehensive in silico study, MET known has possibilities of utilizing as a combination drug with existing antibiotics for TB therapy.¹⁰ and by an extensive in vitro study, MET was reported controlling the growth of drug-resistant *M. tuberculosis* strains via production of mitochondrial reactive oxygen species and facilitates phagosome-lysosome fusion.¹¹ Thus, MET is known as one of highly potential HDT due to target autophagy by AMPK activation or known as mTOR inhibitor.^{11,12}

Moreover, MET is not metabolized by P450 enzymes.^{4,13,14} thus it has no interaction with rifampicin that could decrease the therapy efficacy. However, interaction MET and rifampicin increases the expression of organic cation transporter (OCT1) and hepatic uptake of metformin, leading to an enhanced glucose-lowering.^{4,5,14} MET is also expected enhanced isoniazid (INH) efficacy due to SOD activity.¹⁵ INH a pro-drug, its activation is requiring an interaction with Kat-G produced by *M. tuberculosis*.^{4,14} Kat-G activation also produces oxidative stress – reactive oxygen species (ROS), namely H₂O₂ and alkyl hydroperoxides. ROS is neutralized by an antioxidant, superoxide dismutase (SOD).^{15,16} It is suspected that SOD contributes to the INH-induced bactericidal effects.

However, the mechanism of immune response changes due to the effects of metformin therapy to enhance the effect of anti-TB was not yet clear. This study, due to clarification aims, we measured the change levels of MAP1LC3B, SOD and IFN- γ levels before and after this observation period and as clinical result, we also evaluated the smear reversion in DM-TB coinfection patients.

2. Materials and methods

2.1. Study design

This involved type 2 DM-TB coinfection patients of observational clinical study and simple random allocation technique with two groups, MET group and non MET group. The MET group was the group receiving metformin therapy 1000–1500 mg along with insulin and anti-TB during the intensive periods, while non MET group received insulin and anti-TB therapy. Patients were carried out at outpatient ward of Surabaya Paru Hospital, with criteria: (1) patient DM with new case of TB co-infection, whom were given insulin and TB treatment regimens; (2) positive AFB in sputum smear; (3) patient's age was 25–60 years old; (4) has normal liver function and renal function; (5) not in hypoxia condition, presenting by peripheral oxygen saturation level must be higher than 92%.

The levels of MAP1LC3B, SOD and IFN- γ was measured before and after this observation period and as clinical result, we also evaluated the smear reversion in DM-TB coinfection patients in both of groups.

2.2. Diagnosis and management therapy

The diagnosis of TB was established by (1) clinical symptoms and signs of TB, such: chronic productive cough, unintentional weight loss; (2) positive sputum smear of acid-fast bacilli (AFB) by microscopic Ziehl-Neelsen-stained sputum slides; and (3) chest radiographs with suggestive features of TB. Diagnosis of DM was established by fasting and 2 h after meal blood glucose. HbA1c was measured after 2 months MET therapy, as evaluation.

Patients diagnosed with TB were registered and treated with anti-TB regimens for a period of 6 months in accordance to WHO guidelines.^{17,18} Management therapy for achieving good glycaemic control was insulin therapy. These following drugs were used: MET (Metformin^(R)), insulin (Humulin^(R)), rifampicin (RIF), isoniazid (INH), pyrazinamide (PYR), ethambutol (ETH). MET were given 1000 – 1500 mg in divided daily dose for at least two months or during intensive phase of anti-TB.

2.3. AFB smears

Sputum smears were stained by the Ziehl-Nielsen technique and examined by light microscopy for AFB. Sputum were collected two times: (1) before treatment in order to diagnose and (2) after intensive phase of anti-TB treatment in order to evaluate the treatment.

2.4. Cells culture and ELISA

2.4.1. Cells

MC was obtained from patients' whole blood and 1×10^6 were cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum. Supernatant were harvested after 72 h and prepare for ELISA methods in order to measure the levels of MAP1LC3B, SOD and IFN- γ .

2.4.2. ELISA

MAP1LC3B (MyBioSource MBS760738); SOD (Biovision K335-100); and IFN- γ (RnD DIF50) were used as measurement kits.

3. Result and discussion

3.1. Characteristic of patients

This study's ethical clearance was approved by ethical committee of Surabaya Paru Hospital with no. 09.01/KERS/102.6/2016. During this study period, there were 47 cases of new TB infection and 156 cases (~30%) of that were type 2 DM newly TB co-infection. 42 patients were eligible participated in this observational studies. All the basic conditions in both groups were homogenous ($p > 0.05$) (as seen in Table 1).

In order to prevent MET associated lacto acidosis (MALA) during MET therapy in this study, all patients has been determined at the precondition criterias as seen in Section 2.1. Moreover, there was no incidence of lactic acidosis event during this observation period.¹⁹

3.2. Acid fast bacilli smears

Sputum for AFB smears were collected two times: (1) before treatment; and (2) after intensive phase of anti-TB therapy in order to evaluate the treatment (Table 2).

Data in Table 2 shows that prior to intensive phase of anti-TB therapy none of subjects were having negative AFB smear in both groups. The highest number of AFB count (+3) in MET group was 40.9% and in non MET group was 35%. After 2 months MET therapy accompanying with insulin and anti-TB regimens, 100% of patient of MET group were AFB smear reversion (negative AFB smears result), while only 75% of non MET group had AFB smear reversion. Using the Fisher's exact test, results of different test $p = 0.046$ ($p < 0.005$), which means there is significantly difference of AFB smears reversion between the MET group and the Non MET group.

3.3. Level of MAP1LC3B, SOD, IFN- γ

3.3.1. Autophagy

Autophagy is a fundamental process of cell biology intimately involved in the interaction between *M. tuberculosis* and the phagocytes, including macrophages, dendritic cells (DC) and neutrophils. *M. tuberculosis* has capacity to evade the autophagy of mononuclear phagocytes (MPs) and leverage the intracellular environment as a replication site.^{20,21} Combined with hyperglycemia state and high free radicals of oxygen or nitrogen, infected MPs of DM patients are faced with a pathogen surviving in phagosomes that fail to incorporate the molecular machinery and also fail to fuse with lysosomes to expose bacilli, then, resulting the failure of anti-TB therapy.^{22,23}

Autophagy in TB infection is a combined response of innate and adaptive host immune systems that are essential for the process of *M. tuberculosis* elimination. Microtubule-associated protein 1 light chain 3 (MAP1LC3B/ATG8) is ubiquitin-like modifier that represents autophago-lysosome fusion or better known as autophagy.²⁴ thus in this study MAP1LC3B was used to represent autophagy activity.

MET is known as mTOR inhibitor, by the activation of AMPK.^{9,12,25} Recent researches of MET that has been done in silico, in vitro and animal's in vivo, expressed that MET is a potential adjunct for anti-TB therapy.^{10,12,25}

Table 3 shows that MAP1LC3B level before treatment between MET group and non MET group were alike ($p > 0.005$), thus it shows that patients in both groups, before treatment, were in similar stage of MAP1LC3B. The differences before and after observation period was significant in MET group ($p < 0.005$) while in non MET group was not. Moreover, compared to non MET group, the change of MAP1LC3B after MET therapy during intensive period of anti-TB and insulin was also significant differences. Thus, we concluded that MET improves autophagy and it results 100% AFB smear reversion rate in MET group (as seen in Table 2).


Table 1 – Characteristic of type 2 DM-TB coinfection during observation period of study.

Parameters	MET group	Non MET group	p (difference)
Ages (years old)	44.59 \pm 8.64	48.40 \pm 8.17	0.863
HbA1c (g/dL) (%)	8.82 \pm 1.91	9.52 \pm 2.02	0.379
Oxygen saturation (SpO ₂) (%)	98.06 \pm 0.73	97.47 \pm 0.83	0.308
BUN (mg/dL)	0.95 \pm 0.16	0.93 \pm 0.13	0.980
Creatinine serum (U/L)	23.92 \pm 11.92	27.3 \pm 12.01	0.103
SGOT (U/L)	17.63 \pm 6.16	14.44 \pm 6.48	0.354
SGPT (U/L)	19.22 \pm 8.73	16.09 \pm 7.56	0.509

Table 2 – Acid fast bacilli smears result of type 2 DM-TB patients before and after observation period.

AFB result	MET group (N %)		Non MET group (N %)	
	Before	After	Before	After
Negative	0 (0%)	22 (100%)	0 (0%)	15 (75%)
Scanty/+1	9 (40.9%)	0 (0%)	6 (30%)	5 (25%)
+2	4 (18.2%)	0 (0%)	7 (35%)	0 (0%)
+3	9 (40.9%)	0 (0%)	7 (35%)	0 (0%)
Total	22 (100%)	22 (100%)	20 (100%)	20 (100%)

Please cite this article in press as: Novita BD, et al. Metformin induced autophagy in diabetes mellitus – Tuberculosis co-infection patients: A case study, *Indian J Tuberc.* (2018), <https://doi.org/10.1016/j.ijtb.2018.04.003>

Table 3 – Microtubule-associated Protein1 Light Chain 3B (MAP1LC3B) level of type 2 DM-B patient before and after observation period.




	MAP1LC3B level (pg/mL)  MET group	Non MET group	Between groups differences
	$\bar{x} \pm SD$	$\bar{x} \pm SD$	
Before	1247.7 \pm 551.5	1571.6 \pm 477.7	0.062
After	1859.4 \pm 47.3	1343.6 \pm 607.9	0.004
Before and after differences	$p = 0.000$	$p = 0.830$	

Table 4 – Superoxide dismutase (SOD) level of type 2 DM-B patient before and after observation period.

	SOD's level (U/mL)  MET group	Non MET group	Between groups differences
	$\bar{x} \pm SD$	$\bar{x} \pm SD$	
Before	47.63 \pm 25.76	86.23 \pm 18.63	0.427
After	57.33 \pm 27.34	61.73 \pm 22.57	0.138
Before and after differences	$p = 0.000$	$p = 0.343$	

Table 5 – Interferon (IFN)- γ level of type 2 DM-B patient before and after observation period.


	IFN- γ level (pg/mL)  MET group	Non MET group	Between groups differences
	$\bar{x} \pm SD$	$\bar{x} \pm SD$	
Before	1116.4 \pm 650.2	1430.9 \pm 680	0.072
After	1857.4 \pm 48.77	1313.7 \pm 617.6	0.004
Before and after differences	$p = 0.000$	$p = 0.899$	

Table 6 – Interaction between metformin, level of SOD, MAP1LC3B, IFN- γ , and AFB smears reversion.

No.	Variable	Beta (β)	p
1.	Metformin \rightarrow IFN- γ	0.621	0.000 ^a
2.	Metformin \rightarrow SOD	0.681	0.000 ^a
3.	Metformin \rightarrow MAP1LC3B	0.636	0.000 ^a
4.	Metformin \rightarrow AFB reversion	0.386	0.012 ^a
5.	IFN- γ \rightarrow MAP1LC3B	0.875	0.000 ^a
6.	SOD \rightarrow MAP1LC3B	0.441	0.004 ^a
7.	IFN- γ \rightarrow AFB reversion	0.295	0.015 ^a
8.	SOD \rightarrow AFB reversion	0.267	0.122
9.	MAP1LC3B \rightarrow AFB reversion	0.303	0.021 ^a

Beta (β): regression coefficient, shows the relationship between independent variables to dependent variable (increase or decrease, negative value means inhibit).

p: probability significance value, is stated significant; significant when p value < 0.05.

\rightarrow : affect to.

^a Meaningful.

3.3.2. SOD

SOD contributes to the INH's mycobactericid effect and inducing autophagy.^{4,26}

Table 4 shows that SOD level before treatment between MET group and non MET group were alike ($p > 0.005$), thus it shows that patients in both groups, before treatment, were in similar stage of SOD. The differences before and after observation period was significant in MET group ($p < 0.005$) while in non MET group was not. Moreover, compared to non MET group, determination SOD change as the effect of MET therapy during intensive period of anti-TB was not significant differences in MET group.

Based on those results, we concluded that MET, 1.000–1.500 mg/day, was significantly increased SOD activities which it is enhanced *M. tuberculosis* eliminating process. However, the different of SOD level after observation period in both groups were not significant changes. In our knowledge, SOD activity is generated by the body and adjusted to the existing oxidant levels,¹⁵ thus, this result phenomenon is due to SOD activity triggered not only by MET induced AMPK but also by other factors such oxidative stress, hyperglycemia associated ROS and *M. tuberculosis* elimination process.¹¹

In this study, elevated SOD levels were a synergistic effect of MET and anti-TB therapy. Furthermore, high blood SOD levels in the MET group showed that the oxidative stress in the MET group was also high. The increased oxidative stress in this study was the result of respiratory macrophage cell burst of *M. tuberculosis* phagocytosis process in the blood.^{11,27} The increase of SOD in this study is assumed as high intracellular killing effect of macrophage cell on *M. tuberculosis* and is proven by AFB smear reversion rate in MET group (100% as seen in Table 2).

3.3.3. IFN- γ

IFN- γ is activated by Th1 to maintain IL-12 receptor and provide protection against TB infection.²⁸ Increased of IFN- γ in chronic TB infection is a cellular immune response. Currently, IFN- γ release assay (IGRA) is used as one of the tools of diagnosis of latent TB infection and IFN- γ elevation is one of the indicators of successful treatment in active TB infection.^{29,30} Moreover, IFN- γ plays important key in autophagy.^{21,30}

Table 5 shows that IFN- γ level before treatment between MET group and non MET group were alike ($p > 0.005$), thus it shows that patients in both groups, before treatment, were in similar stage of IFN- γ . The differences before and after observation period was significant in MET group ($p < 0.005$) while in non MET group was not. Moreover, compared to non MET group, IFN- γ elevation as the effect of MET therapy during intensive period of anti-TB was also significant differences. Thus, we concluded that MET improves autophagy and it results 100% AFB smear reversion rate in MET group (as seen in Table 2).

3.4. Interaction between metformin, level of SOD, MAP1LC3B, IFN- γ , and AFB smears reversion

Using regression category, we concluded that AFB smear reversion in this study was influenced by MET through autophagy activity and IFN- γ , while SOD did not affect the AFB smear reversion directly, but through the autophagy (as seen in Table 6). Several studies related to the autophagy in TB infection, suggest that SOD and IFN- γ play important role in autophagy and the results of this study support those information.

4. Conclusion

Research and development of new host-directed therapies aimed at improving clinical outcomes of TB patients, are now beginning to be widely practiced in TB therapy strategies, especially using immunomodulatory drugs. The high number of resistance and failure of therapy in TB infection is one of the reasons for the implementation of the activity. Glucocorticoid and IL-12 drugs are “old” drugs used in new TB treatment strategies.

Some previous studies suggest that MET is an immune-modulator that inhibits cancer cell proliferation stimulator, insulin like growth factor (IGF) and activation of PI3K-AKT. The mechanism of inhibition that is the reason Metformin is used for the prevention and adjuvant therapy for cancer, including breast cancer, ovarian cancer, colorectal cancer and prostate cancer. Thus, several comprehensive studies in silico, in vitro and animal in vivo also suggest that MET may increase the efficacy of anti-TB treatment. However, no human studies have supported it. MET has additive effects on anti-TB due to: (1) MET may increase the expression of organic cation transporter (OCT)-1 rifampicin were instilled in *M. tuberculosis* inhibition of transcription. (2) MET, based on the results of this study and some previous research, could increase the SOD to prevent resistance to isoniazid.

This study provides data on patient clinical changes, the AFB smear reversion in DM-TB coinfection patients (Table 2) so that it can be concluded that MET uses as a insulin combination and has a synergistic effect in enhance anti-TB efficacy. Moreover, MET, based on the results of this study and some previous studies, may be concluded as autophagy inducer or mTOR inhibitor. Nevertheless, the mechanism of action of metformin at the molecular level involving phagosomes and lysosomes remains unknown yet in detail and requires further study. In the even of MET associated

autophagy through the phagosome, it may increase the efficacy of pyrazinamide induced intracellular phagocytosis of *M. tuberculosis*, that need further study.

As preliminary study, this result need to be continued in subsequent studies to develop more effective TB treatment strategies and the development of new adjuvant therapy that works as an immune-modulator by emphasizing the improvement of SOD, autophagy and have less minimal gastrointestinal side effects and the likelihood of more minimal lactic acidosis. As conclusion, MET has the potential of being an additive combination therapy to enhance the bactericidal effect of anti-TB on DM-TB coinfection patients. Metformin enhances the effects of anti-TB and insulin therapy in increasing the AFB smear reversion by increasing autophagy. MET was also relatively safe for DM-TB coinfection patients due to its result in not elevated lactate levels.

Conflicts of interest

The authors have none to declare.

Acknowledgement

The authors would like to thank almighty God for helping us complete this work successfully. We thank Sri Hariastuti and team (Clinical Pathology, Diagnostic Center Dr. Soetomo Hospital, Surabaya, Indonesia) for her technical assistance. We thank Wuryani, Dr., Sp.PD; Kusdiantoro, Dr., Sp.P and Indrayana, Dr., Sp.P as our clinical mentors in Surabaya Paru Hospital. We also thank Prof. Dr., Moh. Amin, Dr., Sp.P(K); Prof. Dr. Jusak, Dr., M.S., Sp.PK(K); Dr. Hari Basuki, Dr., MS. and Prof. Dr. H. Joewono Soerono, Dr., M.Sc., Sp.PD-KR for the kindly discussion.

REFERENCES

- Almeida Da Silva PEA, Palomino JC. Molecular basis and mechanisms of drug resistance in *Mycobacterium tuberculosis*: classical and new drugs. *J Antimicrob Chemother.* 2011;66(7):1417–1430. <http://dx.doi.org/10.1093/jac/ckr173>.
- Ogbera AO, Kapur A, Abdur-Razzaq H, et al. Clinical profile of diabetes mellitus in tuberculosis. *BMJ Open Diabetes Res Care.* 2015;3(1):e000112. <http://dx.doi.org/10.1136/bmjdr-2015-000112>.
- Baker MA, Harries AD, Jeon CY, et al. The impact of diabetes on tuberculosis treatment outcomes: a systematic review. *BMC Med.* 2011;9(81):1–15. <http://dx.doi.org/10.1186/1741-7015-9-81>.
- Brunton L, Chapner B, Knollmann B. In: Brunton L, Chapner B, eds. *The Pharmacological Basis of Therapeutics-Goodman & Gillman-Ed 12th ed.* San Diego, CA: Mc Graw Hill Medical; 2011.
- Thee S, Seddon JA, Donald PR, et al. Pharmacokinetics of isoniazid, rifampin, and pyrazinamide in children younger than two years of age with tuberculosis: evidence for implementation of revised World Health Organization recommendations. *Antimicrob Agents Chemother.* 2011;55(12):5560–5567. <http://dx.doi.org/10.1128/AAC.05429-11>.

6. Clemens DL, Lee B, Xue M, et al. Targeted intracellular delivery of antituberculosis drugs to *Mycobacterium tuberculosis*-infected macrophages via functionalized mesoporous silica nanoparticles. *Antimicrob Agents Chemother*. 2012;56(5):2535–2545. <http://dx.doi.org/10.1128/AAC.06049-11>.
7. Caire-Brändli I, Papadopoulos A, Malaga W, et al. Reversible lipid accumulation and associated division arrest of *Mycobacterium avium* in lipoprotein-induced foamy macrophages may resemble key events during latency and reactivation of tuberculosis. *Infect Immun*. 2014;82(2):476–490. <http://dx.doi.org/10.1128/IAI.01196-13>.
8. Hawn TR, Matheson AI, Maley SN. Host-directed therapeutics for tuberculosis: can we harness the host? *Microbiol Mol Biol Rev*. 2013;77(4):608–627. <http://dx.doi.org/10.1128/MMBR.00032-13>.
9. Wallis RS, Hafner R. Advancing host-directed therapy for tuberculosis. *Nat Rev Immunol*. 2015;15(4):255–263. <http://dx.doi.org/10.1038/nri3813>.
10. Vashisht R, Brahmachari SK. Metformin as a potential combination therapy with existing front-line antibiotics for Tuberculosis. *J Transl Med*. 2015;13(83):1–3. <http://dx.doi.org/10.1186/s12967-015-0443-y>.
11. Singhal A, Jie L, Kumar P, et al. Metformin as adjunct antituberculosis therapy. *Sci Transl Med*. 2014;6(263):159–263. <http://dx.doi.org/10.1126/scitranslmed.3009885>.
12. Restrepo BI. Metformin: candidate host-directed therapy for tuberculosis in diabetes and non-diabetes patients. *Tuberculosis*. 2016;101:S69–S72. <http://dx.doi.org/10.1016/j.tube.2016.09.008>.
13. Madiraju AK, Erion DM, Rahimi Y, et al. Metformin suppresses gluconeogenesis by inhibiting mitochondrial glycerophosphate dehydrogenase. *Nature*. 2014;510(7506):542–546. <http://dx.doi.org/10.1038/nature13270>.
14. Katzung BG, Masters SB, Trevor AJ. *Basic & Clinical Pharmacology*. Twelfth. Singapore: Mc Graw Hill Education (Asia); 2012.
15. Palanisamy N, Manian S. Protective effects of *Asparagus racemosus* on oxidative damage in isoniazid-induced hepatotoxic rats: an in vivo study. *Toxicol Ind Health*. 2012;28(3):238–244. <http://dx.doi.org/10.1177/0748233711410911>.
16. Hofmann-Thiel S, van Ingen J, Feldmann K, et al. Mechanisms of heteroresistance to isoniazid and rifampin of *Mycobacterium tuberculosis* in Tashkent, Uzbekistan. *Eur Respir J*. 2009;33(2):368–374. <http://dx.doi.org/10.1183/09031936.00089808>.
17. Menzies D, Sterling TR. Treatment of *Mycobacterium tuberculosis* infection: time to get a move on? *Ann Intern Med*. 2014;161(6):449. <http://dx.doi.org/10.7326/M14-1719>.
18. van Deun A, Monedero I, Rieder HL, et al. In: Caminero J, ed. *Guidelines for Clinical and Operational Management of Drug-Resistant Tuberculosis* 2013. 2013.
19. Novita BD, Pranoto A, Wuryani. Soediono EI, Mertaniasih NM. A case risk-study of lactic acidosis risk in metformin use in type 2 diabetes mellitus tuberculosis co-infection patients. *Indian J Tuberc*. 2017. <http://dx.doi.org/10.1016/j.ijtb.2017.05.008>.
20. Silva Miranda M, Breiman A, Allain S, Deknuydt F, Altare F. The tuberculous granuloma: an unsuccessful host defence mechanism providing a safety shelter for the bacteria? *Clin Dev Immunol*. 2012;2012:139127. <http://dx.doi.org/10.1155/2012/139127>.
21. Andrew M, Hardy K. Cell death and autophagy in TB. *Semin Immunol*. 2015;26(6):497–511. <http://dx.doi.org/10.1016/j.smim.2014.10.001>.
22. Harries AD, Satyanarayana S, Kumar AMV, et al. Epidemiology and interaction of diabetes mellitus and tuberculosis and challenges for care: a review of diabetes mellitus. *Public Heal Action*. 2013;(May):S3–S9.
23. Sali N, Dembélé SM, Villani P, et al. Systemic exposure to rifampicin in patients with tuberculosis and advanced HIV disease during highly active antiretroviral therapy in Burkina Faso. *J Antimicrob Chemother*. 2012;67(2):469–472. <http://dx.doi.org/10.1093/jac/ckr445>.
24. Ogawa M, Mimuro H, Yoshikawa Y, Ashida H, Sasakawa C. Manipulation of autophagy by bacteria for their own benefit. *Microbiol Immunol*. 2011;459–471. <http://dx.doi.org/10.1111/j.1348-0421.2011.00343.x>.
25. Singhal A, Singhal A, Jie L, et al. Metformin as adjunct antituberculosis therapy. *Sci Transl Med*. 2014;6(263):1–10. <http://dx.doi.org/10.1126/scitranslmed.3009885>.
26. Singhal J, Agrawal N, Vashishta M, et al. Suppression of dendritic cell-mediated responses by genes in calcium and cysteine protease pathways during *Mycobacterium tuberculosis* infection. *J Biol Chem*. 2012;287(14):11108–11121. <http://dx.doi.org/10.1074/jbc.M111.300319>.
27. Keinath NF, Kierszniowska S, Lorek J, et al. PAMP (pathogen-associated molecular pattern)-induced changes in plasma membrane compartmentalization reveal novel components of plant immunity. *J Biol Chem*. 2010;285(50):39140–39149. <http://dx.doi.org/10.1074/jbc.M110.160531>.
28. Abbas AK, Lichtman A. In: Abbas AK, Lichtman A, Pillai S, eds. *Cellular and Molecular Immunology* 7th ed. Philadelphia, PA: Saunders; 2012.
29. Chee CBE, KhinMar KW, Gan SH, et al. Tuberculosis treatment effect on T-cell interferon-gamma responses to *Mycobacterium tuberculosis*-specific antigens. *Eur Respir J*. 2010;36(2):355–361. <http://dx.doi.org/10.1183/09031936.00089808>.
30. Matsushita I, Hang NTL, Hong LT, et al. Dynamics of immune parameters during the treatment of active tuberculosis showing negative interferon gamma response at the time of diagnosis. *Int J Infect Dis*. 2015;40:39–44. <http://dx.doi.org/10.1016/j.ijid.2015.09.021>.
31. Bento CF, Empadinhas N, Mendes V. Autophagy in the fight against tuberculosis. *DNA Cell Biol*. 2015;34(4):228–242. <http://dx.doi.org/10.1089/dna.2014.2745>.
32. Devenish RJ, Lai S. Autophagy and burkholderia. *Immunol Cell Biol*. 2015;93(1):18–24. <http://dx.doi.org/10.1038/icb.2014.87>.
33. Melkam W, Gebremedhin H, Abirha S, Masresha B, Molla F. Glucocorticosteroids: as adjuvant therapy for bacterial infections. *Int J Pharm Sci Res*. 2015;6(1):145–151.
34. Kourelis TV, Siegel RD. Metformin and cancer: new applications for an old drug. *Med Oncol*. 2012;29(2):1314–1327. <http://dx.doi.org/10.1007/s12032-011-9846-7>.
35. Kasznicki J, Sliwinski A, Drzewoski J. Metformin in cancer prevention and therapy. *Ann Transl Med*. 2014;2(6):1–11. <http://dx.doi.org/10.3978/j.issn.2305-5839.2014.06.01>.
36. Almeida VD. Revisiting anti-tuberculosis activity of pyrazinamide in mice. *Mycobact Dis*. 2014;4(2):2–7. <http://dx.doi.org/10.4172/2161-1068.1000145>.

Metformin induced autophagy in diabetes mellitus – Tuberculosis co-infection patients: A case study

ORIGINALITY REPORT

% **14**
SIMILARITY INDEX

% **10**
INTERNET SOURCES

% **5**
PUBLICATIONS

% **0**
STUDENT PAPERS

PRIMARY SOURCES

1 repository.unair.ac.id % **5**
Internet Source

2 www.scitechnol.com % **2**
Internet Source

3 Andrew H. Moraco, Hardy Kornfeld. "Cell death and autophagy in tuberculosis", Seminars in Immunology, 2014 % **1**
Publication

4 www.hts.org.gr % **1**
Internet Source

5 Prema Nedungadi, Akshay Jayakumar, Raghu Raman. "Personalized Health Monitoring System for Managing Well-Being in Rural Areas", Journal of Medical Systems, 2017 % **1**
Publication

6 Y. Ma, Y. Pang, W. Shu, Y.-H. Liu, Q.-P. Ge, J. Du, L. Li, W.-W. Gao. "Metformin reduces the relapse rate of tuberculosis patients with % **1**

diabetes mellitus: experiences from 3-year follow-up", European Journal of Clinical Microbiology & Infectious Diseases, 2018

Publication

-
- | | | |
|---|--|---------------|
| <div style="background-color: #800000; color: white; display: inline-block; width: 30px; height: 30px; text-align: center; line-height: 30px;">7</div> | <p>Restrepo, Blanca I.. "Metformin: Candidate host-directed therapy for tuberculosis in diabetes and non-diabetes patients", Tuberculosis, 2016.</p> | <p>%1</p> |
| <hr/> | | |
| <div style="background-color: #003366; color: white; display: inline-block; width: 30px; height: 30px; text-align: center; line-height: 30px;">8</div> | <p>Denny John, Prabir Chatterjee, Shruti Murthy, Ramesh Bhat, Baba Mayaki Musa. "Cost effectiveness of decentralised care model for managing MDR-TB in India", Indian Journal of Tuberculosis, 2017</p> | <p><%1</p> |
| <hr/> | | |
| <div style="background-color: #800080; color: white; display: inline-block; width: 30px; height: 30px; text-align: center; line-height: 30px;">9</div> | <p>docplayer.net</p> | <p><%1</p> |
| <hr/> | | |
| <div style="background-color: #006633; color: white; display: inline-block; width: 30px; height: 30px; text-align: center; line-height: 30px;">10</div> | <p>scholarworks.waldenu.edu</p> | <p><%1</p> |
| <hr/> | | |
| <div style="background-color: #000080; color: white; display: inline-block; width: 30px; height: 30px; text-align: center; line-height: 30px;">11</div> | <p>Lee, Heung-Man, Il-Ho Park, Jae-Min Shin, Hyun-Sun Yoon, Gyeong Yul Park, Margit Zeher, Katsuhiko Matsui, Saki Tamai, Reiko Ikeda, Drsushil Suri, Dranu Suri, Marzieh Heidarzadeh Arani, Azwin Lubis, Anang Endaryanto, Shinichiro Koga, Lee Ju Suk, Yasunobu Tsuzuki, Seo Hyeong Kim, Jung U. Shin, Ji Yeon Noh, Shan Jin, Shan Jin, Hemin</p> | <p><%1</p> |

Lee, Jungsoo Lee, Chang Ook Park, Kwang Hoon Lee, Kwang Hoon Lee, Fatma Merve Tepetam, Chun Wook Park, Jee Hee Son, Soo Ick Cho, Yong Se Cho, Yun Sun Byun, Yoon Seok Yang, Bo Young Chung, Hye One Kim, Hee Jin Cho, Yoshinori Katada, Toshio Tanaka, Akihiko Nakabayashi, Koji Nishida, Kenichi Aoyagi, Yuki Tsukamoto, Kazushi Konma, Motoo Matsuura, Jung-Won Park, Yoshinori Harada, Kyoung Yong Jeong, Akiko Yura, Maiko Yoshimura, Tae-Suk Kyung, Young Hyo Kim, Chang-Shin Park, Tae Young Jang, Min-Jeong Heo, Ah-Yeoun Jung, Seung-Chan Yang, Hye One Kim, Yong Se Cho, Yun Sun Byun, Yoon Seok Yang, Bo Young Chung, Jee Hee Son, Chun Wook Park, Hee Jin Cho, Oliver Pfaar, Angelika Sager et al. "XXIV World Allergy Congress 2015 : Seoul, Korea. 14-17 October 2015", World Allergy Organization Journal, 2016.

Publication

12

www.researchgate.net

Internet Source

<% 1

13

Antony, Cecil, Subhash Mehto, Brijendra K. Tiwari, Yogendra Singh, and Krishnamurthy Natarajan. "Regulation of L-type Voltage Gated Calcium Channel CACNA1S in Macrophages upon Mycobacterium tuberculosis Infection",

<% 1

EXCLUDE QUOTES ON

EXCLUDE ON
BIBLIOGRAPHY

EXCLUDE MATCHES < 10
WORDS