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biortech

5Bio-oil from cassava peel: A potential renewable energy source Ong Lu Ki a,1, Alfin Kurniawan a,1, Chun Xiang Lin b, Yi-Hsu Ju c, Suryadi

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8highlights" Cassava peel was utilised as a potential biomass feedstock to produce bio-oil. "A relatively high yield of bio-oil was obtained through slow pyrolysis at 525 °C." The distribution of organic moieties in bio-oil is not dependent on temperature. "The physicochemical

properties of bio-oil satisfied ASTM D7544 requirements. " The yield of pyrolysis products was predicted by kinetic-based mechanistic model. article info Article history: Available online 9 February 2013 Keywords: Cassava peel Bio-oil Fixed-bed reactor Slow pyrolysis Mechanistic model abstract In this work, liquid biofuel (bio-oil) was produced by pyrolizing cassava peel. The experiments were con- ducted isothermally in a fixedbed tubular reactor at temperatures ranging from 400 to 600 °C with a heating rate of 20 °C/min. The chemical compositions of bio-oil were analyzed by a gas chromatography mass spectrometry (GC-MS) technique. For the optimization of liquid product, temperature was plotted to be the most decisive factor. The maximum yield of bio-oil ca. 51.2% was obtained at 525 °C and the biofuel has a gross calorific value of 27.43 MJ/kg. The kinetic-based mechanistic model fitted well with experimental yield of pyrolysis products with the mean squared error (MSE) of 13.37 (R2 = 0.96) for solid (char), 16.24 (R2 = 0.95) for liquid (bio-oil), and 0.49 (R2 = 0.99) for gas. Ó 2013 Elsevier Ltd. All rights reserved. 1. Introduction Current technologies available to convert biomass into fuels can be classified into three categories based on their methodologies namely biochemical, thermal, and thermochemical conversion. Thermochemical conversion of biomass includes a number of pro- cesses such as liquefaction, gasification, and pyrolysis (Park et al., 2012). Among these, pyrolysis is the main spotlight of thermo- chemical conversion of biomass that produces solid and liquid fuels, both are easy to handle and transport (Van de Velden et al., 2010). The liquid fuel (bio-oil) contains various added-value chem- icals in economical recoverable concentrations and can be up- graded into refined fuels for transportation purpose (Volli and Singh, 2012). Cassava (Manihot esculenta) is extensively cultivated in Indone- sia as one of major staple food resources as well as a multipurpose agroindustrial crop. Cassava starch is a high value commodity for ↑ Corresponding author. Tel.: +62 313891264; fax: +62 313891267. E-mail address: suryadiismadji@yahoo.com (S. Ismadji). 1 These authors contributed equally to this work. 0960-8524/\$ - see front matter Ó 2013 Elsevier Ltd. All rights reserved.

http://dx.doi.org/10.1016/j.biortech.2013.01.122 various industries such as brewing, paper, adhesives, plastics, and chemicals. The peel waste that generated from the cassava starch manufacturing industries is of great concern to the environment. Since cassava peel contains cyanogenic glucosides mainly linama- rin and lotaustralin, the disposal and improper handling of this waste often create an environmental problem due to the release of hydrogen cyanide (HCN) from cyanogenesis activity of endoge- nous linamarase enzyme (Cooke, 1978). Several research groups have employed cassava peel as a potential feedstock to prepare electrode materials for supercapacitors (Ismanto et al., 2010) and cheap biosorbent for sequestering heavy metal ions (Kosasih et al., 2010; Kurniawan et al., 2011). Several studies on the utilisation of cassava plant residues for bio-oil production have recently been reported in the literatures (Pattiya, 2011; Pattiya et al., 2012). In their studies, Pattiya and his co-workers use rhizome and stalk of cassava plant as biomass feedstocks to obtain bio-oil by fast pyrolysis process in two types of reactors namely fluidized-bed reactor (Pattiya, 2011) and free- fall reactor (Pattiya et al., 2012). Another work has been conducted by Weerachanchai et al. (2011) dealing with slow paralysis of cassava pulp residue and palm kernel cake and its product 158 O.L. Ki et al. / Bioresource Technology 145 (2013) 157-161 characterization includes solid, liquid, and gas. In this work, cassava peel is used as a raw material to produce bio-oil through slow pyrolysis process in a fixed-bed reactor. The effect of pyrolysis temperatures on the physicochemical properties of bio-oil was studied. Furthermore, a kinetic-based mechanistic model for predicting the yield of char, bio-oil, and gas was also given in this paper. 2. Methods 2.1. Preparation and analysis of biomass feedstock Cassava peel was collected from a cassava starch factory located near Surabaya, Indonesia. Prior to use, the biomass was repeatedly washed using tap water to remove surface dirt and dried in an oven for 24 h at 110 °C to substantially reduce its moisture content from about 10% to less than 2%. Subsequently, the dried peel was crushed by an IKA-Labortechnick grinder and sieved pass through 80/100 mesh screen to obtain powder with particle sizes of 150–180 lm. The chemical composition of the biomass was analyzed by a Perkin-Elmer 2400 CHNS/O elemental-analyzer and the following results were obtained: C of 53.7%, H of 7.1%, N of 1.2%, S of 0.1%, and O of 37.9% (measured by difference). The proximate analysis of cassava peel was conducted according to the established proce- dures of ASTM D4442-07 for moisture content, ASTM E1755-01 for ash content, and ASTM D3175-11 for volatile matter content. The fixed carbon content was calculated by subtracting the initial weight of the sample (100%) by total percentage of moisture, ash, and volatile matter. The proximate analysis results of cassava peel were given as follows: fixed carbon (FC) of 21.4%, volatile matter (VM) of 74.5%, ash (A) of 2.3%, and moisture (M) of 1.8%. The heat- ing value of the biomass was 17.9 MJ/kg (reported as higher heat- ing value or gross caloric value), as measured by an oxygen bomb calorimeter (Model 1341, Parr Instrument Co.) by following ASTM D5865–12 test method. 2.2. Thermogravimetric analysis (TGA) Thermal stability of the biomass material was studied by ther- mogravimetric analysis. The analysis was performed in a TGA/ DSC 1 STARO System (Mettler-Toledo) with a heating and cooling rate of 10 °C/min and nitrogen (99.99%) was used as the carrier gas at a constant flow rate of 70 ml/min. Approximately, 10 mg of sample was heated in nitrogen atmosphere from room temper- ature to 800 °C at a given heating rate. 2.2. Fixed-bed pyrolysis experiments Slow pyrolysis experiments of cassava peel were performed in a fixed-bed reactor system. The reactor was made from 316-grade stainless steel with a maximum working temperature of 1000 °C at atmospheric pressure. Brief procedure of the experiments is de-scribed as follows: One hundred grams of cassava peel were intro- duced into the reactor. Subsequently, nitrogen at a flow rate of 3 L/ min was introduced to the reactor for removing air and maintain- ing inert condition. The reactor was heated at 20 °C/min from room temperature to desired temperatures (400-600 °C) and held iso- thermally for a certain time. After pyrolysis time reached, the reac- tor was cooled rapidly to room temperature in order to avoid any advancing reactions. The gas and vapor products that generated from the thermal cracking of biomass were passed to a condenser in which the condensable volatiles were converted to liquid form and collected in a measuring

cylinder as bio-oil. At the exit of condensing system, a cotton wool filter was installed to capture low partial pressure hydrocarbons. The non-condensable gases to- gether with the carrier gas were passed to a flowmeter where the volume of exit gases was recorded. The yield of char and bio-oil was determined by weighing the mass of both products until con-stant using an analytical balance (Precisa 3000D). 2.3. Analysis of physical and chemical properties of bio-oil The chemical composition of bio-oil was analyzed by a Shimadzu GC/MS QP2010 equipped with a DB-5 MS capillary col- umn (30 m 0.25 mm 0.25 lm) as the stationary phase. The injector temperature was set at 250 °C while the oven temperature was initially programmed at 60 °C for 1 min then ramped at 10 °C/ min to 300 °C and held for 5 min. Helium was employed as the car- rier gas at a flow rate of 1.5 ml/min. For the mass selective detector (MSD), the temperature of the ion source was 230 °C. The sample was injected into the column in split mode with a ratio of 1:30. Identification of the chemical constituents in bio-oil was con-ducted by comparing the mass spectra of the sample with the Na- tional Institute of Standards and Technology (NIST) database. The gross heat of combustion of bio-oil was determined by an oxygen bomb calorimeter based on ASTM D240. Other physical and chem- ical properties were analyzed by following ASTM test methods: density (ASTM 4052), pH (ASTM E70), kinematic viscosity (ASTM D445), pyrolysis solids content (ASTM D7579), water content (ASTM E203), sulfur content (ASTM D4294), ash content (ASTM D482), pour point (ASTM D97), and flash point (ASTM D93), 3. Results and discussion 3.1. Thermal characteristics of the biomass Thermogravimetric analysis was conducted in order to obtain valuable information regarding the thermal stability of the materials, associated with the mass change as a function of temperature and/or time (figure not shown here). At initial heating zone (T < 200 °C), the weight reduction in biomass primarily attributed to the moisture removal (or physically-bound water). The active pyrolysis zone was observed in temperature range of 200–500 °C where the weight of biomass considerably diminished due to the decomposition of natural polymer materials such as hemicellulose, cellulose, and lignin. Thermal degradation of hemicellulose and cellulose took place at temperatures between 200 and 350 °C while lignin was decomposed at higher temperatures, typically between 280 and 500 °C (Park et al., 2012). 3.2. Effect of pyrolysis temperature on the distribution of products The yield percentage of solid, liquid, and gas obtained from slow pyrolysis of cassava peel at various temperatures are shown in Ta- ble 1. It can be seen that the yield of char diminished from 37.5% to 20.8% while the yield of gases continuously rise from 23.8% to 37.6% with increasing temperatures from 400 to 600 °C. This might be ascribed to the enhanced extent of primary cracking reaction of the biomass at higher temperatures. At temperatures above 525 °C, the combination of primary and secondary crackings increases the yield of gases. In other hand, the experimental trend of liquid yield with temperature was slightly different to that of solid and gas in which the yield of bio-oil increases and reaches its maximum at 525 °C then it gradually decreased from 51.2% to 41.6% at higher temperature, likely due to the secondary thermal cracking. O.L. Ki et al. / Bioresource Technology 145 (2013) 157-161 159 Table 1 Yield and products distribution from slow pyrolysis process of cassava peel. T (°C) Experimental Simulation Solid (wt.%) Liquid (wt.%) Gas (wt.%) Solid (wt.%) Liquid (wt.%) Gas (wt.%) 400 37.5 425 34.3 450 29.6 475 26.8 500 25.1 525 24.3 550 22.9 575 21.4 600 20.8 Mean squared error (MSE) Coefficient of determination (R2) 38.7 23.8 41.8 23.9 46.9 23.5 48.2 25.0 50.3 24.6 51.2 24.5 48.7 28.4 45.4 33.2 41.6 37.6 41.3 37.9 32.7 29.5 27.6 26.7 25.4 23.2 22.9 34.6 38.0 43.0 46.0 47.8 47.5 47.4 43.6 43.4 13.37 0.96 16.24 0.95 24.1 24.2 24.4 24.5 24.6 25.7 27.4 32.8 33.7 0.49 0.99 3.3. Kinetic-based mechanistic modeling of experimental yield of pyrolysis products The kinetic-based mechanistic model was purposely developed in this study for predicting the yield of pyrolysis products at any temperature and time. The reaction scheme of slow pyrolysis pro- cess in this study was similar to that of Van de Velden et al. (2008). Based on this reaction scheme, the kinetic equations representing the decomposition rate of biomass and the formation rate of char, liquid, and gas as a function of time can be expressed as follows: dXB ok1 b k2 b k3b XB ¼¼ k XB o1b dt dXdtG1 ¼ k1 X B ð2Þ dXO1 dt ¼ k2 XB ð3Þ dXC ¼ k3 X B ð4Þ dt dXO2 dXG2 ¼ k4 XO1 ¼ ð5Þ dt dt where X is the

mass fraction and the subscripted notations B, C, G1, O1, G2, and O2 refer to biomass, char, primary gases, primary liquid, secondary gases, and secondary liquid, respectively. The time do- main (t) was designated as a time when the heating was started un-til the process completed. Concerning the real reaction system in this work, the residence time of biomass in the reactor where pri- mary cracking took place was considered as the whole time domain. At initial condition (t = 0), XC = XG1 = XO1 = XG2 = XO2 = 0 and XB = 1. Integrating Eqs. (1)–(5) from t = 0 to t = t gives: XB 1/4 expð ktÞ ð6Þ XG1 1/4 k1 k 1 expð ktÞ ð7Þ XO1 ¼ k2 k 1 expð ktÞ ð8Þ XC ¼ k3 k 1 expð ktÞ ð9Þ The yield of secondary liquid (O2) and secondary gases (G2) on each time interval of the hydraulic time (i.e. the volume of the reactor divided by the N2 flow rate) was simultaneously deter- mined along with the yield of primary oil by applying Euler numer- ical method on Eq. (5). The first-order rate constants in Eqs. (6)–(9) were determined using the same approach described by Van de Velden et al. (2008). In this regard, the dynamic TGA experiments with a heating rate of ca. 100 K/min was used to obtain an accurate value of the overall rate constant of primary cracking reaction (k, s 1). The average values of rate constants for the formation of pri- mary gases (G1) and secondary products (G2 and O2) were taken from the literature (Di Blasi, 2005). Once the values of k1 (the for- mation rate constant of primary gases, s 1) and k4 (the rate con- stant of secondary cracking, s 1) had been known, the values of remaining rate constants, k2 (the formation rate constant of pri- mary oil, s 1) and k3 (the formation rate constant of char, s 1)), could be determined (Van de Velden et al., 2008). The calculated values of all rate constants were given as follows: k 1/4 28057:18expð 81094:76=RTsÞ k1 1/4 14300expð 106500=RTsÞ k2 1/4 21332:77expð 81053:19=RTsÞ k3 1/4 6431:74expð 81094:76=RTsÞ k4 1/4 7900expð 81000=RTsysÞ ð10Þ ð11Þ ð12Þ ð13Þ ð14Þ In this study, the effect of thermal gradient was taken into ac- count because slow heating rates provide small amounts of heat to the system that gradually increases the temperature inside the biomass particle, unlike the case of fast pyrolysis process with a very high heating rate that able to raise the interior temperature of the particle instantaneously. The temperature distribution along the radius of the biomass particle was described by the Fourier's law of heat conduction that has mathematical form: @T ## @t gtrue:Cp @r2 b r @r ¼ j "@2T 2 @T "@2T 2 @T ¼ D 2 b ð15Þ @r r @r with the following boundary conditions: i @T @r r¼Rp ¼ h ðTs TsysÞ ð16Þ @T @r r¼0 ¼ 0 ð17Þ The surface temperature, Ts (K), could be determined by solving Eq. (15) numerically with Crank-Nicolson implicit scheme. In solv- ing Eq. (14), the temperature change in the system during heating period and the particle shrinkage were included. The initial tem- perature of the particles was assumed to be uniform at room tem- perature (30 °C) in any radius position. For simplification, several assumptions were applied in developing the model: (i)

9the shape of the biomass particle is spherical. (ii) the mechanism of heat transfer inside the biomass particle

is only conduction. (iii) the temperature gradient inside the biomass particle is a function of the radius position with surface temperatures less equal to the temperature in the system (Ts 6 Tsys). (iv) Some physical properties of the biomass

12such as density, specific heat capacity, and thermal conductivity are constant. The set of

parameter values used in the calculation is as follows: h = 5.69 + 0.0098 Tsys (Ahuja et al., 1996), Cp = 1449 J kg 1 K 1, j = 0.156 W m 1 K 1, qbulk = 289 kg m 3, t = 0.226, and D = 160 O.L. Ki et al. / Bioresource Technology 145 (2013) 157-161 55 50 45 Yield, % 40 35 30 25 20 15 350 400 450 500 550 600 650 Temperature, oC Fig. 1. Comparison results between simulated (solid lines) and experimental yield of pyrolysis products (d solid, j liquid, gas). 2.8834 10 7 m2.s 1. The simulation process was run by using computer programming on the MATLAB platform (Version 7.11, Mathworks Inc.). Thermal conductivity (j, W m 1 K 1),

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13specific heat capacity (Cp, J kg 1 K 1), bulk density (qbulk, kg m 3),
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and void fraction (t) of the biomass particle are experimentally determined based on ASTM E1952, ASTM E1269, and ASTM C29, respectively. True density of the biomass particle was calculated by using the following equation: qtrue 1/4 ð1 q bulk tÞ ð18Þ Fig. 1 shows the comparison results between experimental (so- lid symbols) and simulated (solid lines) yield of solid, liquid, and gas products. A slight error between simulation and experimental results, as reflected by the coefficient of determination (R2) values, might be ascribed to several following factors such as the non-ide- ality of the interparticle heat transfer in the bed, the neglected pos- sible carbon-steam and/or carbon-carbon dioxide reactions, and the resistance of carrier gas flow in the pores of the solid particle and void fraction of the bed. 3.4.

10Physical and chemical properties of bio-oil The organic fraction in bio-oil

comprises of acids, esters, alde- hydes, ketones, phenols, alcohols, hydrocarbons, and some uniden- tified compounds. The detail composition of organic compounds in bio-oil is summarized in Table 2. As shown in Table 2, there is no a

14Table 3 The physicochemical properties of resulted bio- oil. Properties Biooil

ASTM D7544

15Gross calorific value (MJ/kg) pH Kinematic viscosity (cSt) at 40

°C Density (kg/dm3) at 20 °C Pyrolysis solids content (% wt.) Water content (% wt.)

4Sulfur content (% wt.) Ash content (% wt.) Flash point (°C) Pour point (°C)

27.43 4.26 35.8 1.27 0.12 23.74 0.01 0.08 78 2 15 min -

4125 max 1.1-1.3 2.5 max 30 max

40.05 max 0.25 max 45 min 9 max

definite trend on the organic distributions in bio-oil as a function of temperature, considering the reaction complexity in the pyrolysis process. Several major components of acid groups in bio-oil were acetic acid, formic acid, propionic acid, and butyric acid. For alcohol groups, methanol is predominant while ethanol was found in a small amount. Phenolic compounds in bio-oil might be generated from the pyrolysis of lignin (Bertero et al., 2012). The

11physicochemical properties of resultant bio-oil are given in Table 3. Several properties of bio-oil

2such as viscosity, density, and flash point

are known to be the crucial characteristics for combus- tion purpose in some equipments such as boiler, furnace, gas tur- bines, and engines (Volli and Singh, 2012). The flash point of resultant bio-oil was 1.73-fold higher than that of ASTM require- ment (78 °C vs. 45 °C), indicating safer handling and storage of this liquid fuel in high temperature environment. However, the direct use of this fuel for combustion purpose in the conventional diesel engines is not recommended since it contains many kinds of con-taminants and has a much higher viscosity of ca. 35.8 cSt than commercial diesel fuels of ca. 1.3–5.5 cSt (ASTM D975). High vis- cosity led

2to poor atomization and an incomplete combustion of the fuel

as well as the

2formation of excessive carbon deposits on the injection nozzles and the combustion chamber (Volli and

Singh, 2012). Another issue to be tackled was

2high oxygen content in bio-oil that

affected chemical and thermal stability, corrosivity, and immiscibility with hydrocarbon fuels. For upgrading purpose, several methods are available such as catalytic hydrotreatment to lower the oxygen content (Elliott and Neuenschwander, 1996), supercritical water treatment (SCW) to increase the heating value (Duan and Savage, 2011), or esterification–distillation for removing a large quantity of water (Junming et al., 2008). 4. Conclusions The yield of bio-oil from slow pyrolysis of cassava peel were ranged between 38.7% and 51.2%. The pyrolysis peak temperature Table 2 Chemical composition of bio-oil at various temperatures.

Compounds (wt.%, dry basis) Temperature (°C) 400 425 450 475 500 525 550 575 600 Acids Alcohols Aldehydes and ketones Esters Ethers Phenols Furans Hydrocarbons and its derivatives Nitrogen compounds Oxygenated cyclic compounds Unknowns 20.18 17.91 2.25 3.61 14.32 14.58 8.93 7.46 6.82 5.43 14.89 15.47 6.41 7.52 0.89 1.22 1.21 1.47 3.94 2.98 20.16 22.35 17.65 22.47 2.07 2.98 8.91 12.47 9.22 8.41 5.79 4.14 19.23 10.48 5.63 7.31 1.54 1.21 1.32 1.89 2.71 2.69 25.93 25.95 18.59 19.31 1.96 2.54 17.21 7.93 7.59 7.62 5.64 4.87 17.44 18.03 7.45 6.90 0.83 1.32 1.10 1.18 3.07 3.58 19.12 26.72 24.12 23.22 3.01 2.29 13.74 14.56 8.14 8.88 4.21 3.98 16.88 17.41 6.08 5.41 1.98 2.11 1.48 1.51 3.49 2.55 16.87 18.08 18.94 2.09 12.88 9.01 6.02 18.63 6.98 2.61 1.73 3.41 17.70 O.L. Ki et al. / Bioresource Technology 145 (2013) 157–161 161 that gave maximum liquid yield was observed at 525 °C. The bio- oil was mainly composed of several organic classes such as acids, esters, aldehydes, ketones, phenols, alcohols, hydrocarbons, and other unidentified compounds. All physical and chemical proper- ties of bio-oil satisfied ASTM D7544 standard. The kinetic-based mechanistic model gives good accuracy in representing the exper- imental

5yield of pyrolysis products with the mean squared error of 13.37

for solid (char), 16.24 for liquid (bio-oil), and 0.49 for gas. References Ahuja, P., Kumar, S., Singh, P.C., 1996. A model for primary and heterogeneous secondary reactions of wood pyrolysis. Chem. Eng. Technol. 19, 272–282. Bertero, M., de la Puente, G., Sedran, U., 2012. Fuels from bio-oils: bio-oil production from different residual sources, characterization and thermal conditioning. Fuel 95, 263–271. Cooke, R.D., 1978. An enzymatic assay for the total cyanide content of cassava (Manihot esculenta Crantz), J. Sci. Food Agric. 29, 345–352. Di Blasi, C., 2005. Kinetics and modeling of biomass pyrolysis. In: Bridgwater, A.V. (Ed.), Fast Pyrolysis of Biomass: A Handbook Volume 3. CPL Press, Newbury, pp. 121-146. Duan, P., Savage, P.E., 2011. Upgrading of crude algal bio-oil in supercritical water. Bioresour. Technol. 102, 1899-1906. Elliott, D.C., Neuenschwander, G.G., 1996. Liquid fuels by low-severity hydrotreating of biocrude. In: Bridgwater, A.V., Boocock, D.G.B. (Eds.), Developments in Thermochemical Biomass Conversion, vol. 1. Blackie Academic & Professional, London, pp. 611-621. Junming, X., Jianchun, J., Yunjuan, S., Yanju, L., 2008. Biooil upgrading by means of ethyl ester production in reactive distillation to remove water and to improve storage and fuel characteristics. Biomass Bioenergy 32, 1056–1061. Ismanto, A.E., Wang, S., Soetaredjo, F.E., Ismadji, S., 2010. Preparation of capacitor's electrode from cassava peel waste. Bioresour. Technol. 101, 3534–3540. Kosasih, A.N., Febrianto, J., Sunarso, J., Ju, Y.H., Indraswati, N., Ismadji, S., 2010. Sequestering of Cu(II) from aqueous solution using cassava peel (Manihot esculenta). J. Hazard. Mater. 180, 366–374. Kurniawan, A., Kosasih, A.N., Febrianto, J., Ju, Y.H., Sunarso, J., Indraswati, N., Ismadji, S., 2011. Evaluation of cassava peel waste as low cost biosorbent for Ni- sorption: equilibrium, kinetics, thermodynamics and mechanism. Chem. Eng. J. 172, 158–166. Park, Y.K., Yoo, M.L., Heo, H.S., Lee, H.W., Park, S.H., Jung, S.C., Park, S.S., Seo, S.G., 2012. Wild reed of Suncheon Bay: potential bio-energy source. Renewable Energy 42, 168-172. Pattiya, A., 2011. Bio-oil production via fast pyrolysis of biomass residues from cassava plants in a fluidised-bed reactor. Bioresour. Technol. 102, 1959–1967. Pattiya, A., Sukkasi, S., Goodwin, V., 2012. Fast pyrolysis of sugarcane and cassava residues in a free-fall reactor. Energy 44, 1067–1077. Van de Velden, M., Baeyens, J., Boukis, I., 2008. Modeling CFB biomass pyrolysis reactors. Biomass Bioenergy 31, 128-139. Van de Velden, M., Baeyens, J., Brems, A., Jannsens, B., Dewil, R., 2010. Fundamentals, kinetics and endothermicity of the biomass pyrolysis reaction. Renewable Energy 35, 232-242. Volli, V., Singh, R.K., 2012. Production of bio-oil from de-oiled cakes by thermal pyrolysis. Fuel 96, 579-585. Weerachanchai, P., Tangsathitkulchai, C., Tangsathitkulchai, M., 2011. Characterization of

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