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35 **Chemical Engineering Journal** 231 (2013) 406–413 **Contents lists available at ScienceDirect Chemical Engineering Journal journal homepage: www.elsevier.com/locate/cej**

17 **Recovery of catechin and epicatechin from sago waste effluent: Study of kinetic and binary adsorption isotherm studies Felycia Edi Soetaredjo a, Suryadi Ismadji b, Shella Permatasari Santoso a, Ong Lu Ki b, Alfin Kurniawan**

b,

6 **Yi-Hsu Ju a,† a Department of Chemical Engineering, National Taiwan University of Science and Technology, 43, Sec 4., Keelung Rd., Taipei, Taiwan b Department of Chemical Engineering, Widya Mandala Surabaya Catholic University, Kalijudan 37, Surabaya 60114, Indonesia**

highlights

19 **Recovery of catechin and epicatechin from sago waste effluent.** Supercritical CO₂ desorption **of catechin and**

epicatechin from activated carbon. Adsorption of catechin and epicatechin in binary system.

31 **article info Article history: Received 27 May 2013 Received in revised form 16 July 2013 Accepted 17 July 2013 Available online 26 July 2013 Keywords:**

Catechin Epicatechin Binary adsorption Supercritical CO₂ extraction abstract The recovery of valuable phenolic compounds (catechin and epicatechin) from sago waste effluent by activated carbon adsorption was studied in this work. Single and binary component of catechin and epi- catechin was studied in static mode using synthetic effluent.

12 **Langmuir model was applied to evaluate the adsorption equilibrium data in single system.** In binary system, **the modified extended Langmuir model**

by inclusion of surface coverage was used to represent the adsorption data. All experimental data were well represented either by Langmuir model or its modified form. For kinetic study, it was found that pseudo-second order model represents the experimental data better than pseudo-first order model. Adsorption study using real sago waste effluent indicated that activated carbon adsorption process is promising for recovery of valuable phenolic compounds. Supercritical CO₂ extraction of phenolic compounds from activated carbon resulted in low recovery. The addition of ethanol as co-solvent significantly enhanced the desorption of catechin and epicatechin from activated carbon. Ó 2013 Elsevier B.V. All rights reserved. 1. Introduction Phenolic compounds are universally distributed in plants such as leave, trunk, root, seed, and fruit. They are usually are bounded within plant tissues. Phenolic compounds as antioxidants are believed

33 **to contribute to health benefits through several possible mechanisms** such as **by quenching free radicals, chelating transition metals, reducing peroxides and stimulating antioxidative enzyme defense.**

32 **Natural antioxidants are known to exhibit a wide range of biological effects including anti-bacterial, antiviral, anti-inflammatory, anti-allergic, antithrombotic and vasodilatory**

activities [1].

8The antioxidant activity of phenolic compounds is mainly attributed to their redox properties, which allow them to act as reducing agents, hydrogen donors and quenchers of singlet oxygen. In addition, they may also possess metal chelating properties.

Phenolic compounds possess aromatic structure along with hydroxyl ↑ Corresponding author. Tel.: +886 (2) 2737 6611; fax: +886

18(2) 2737 6644. E-mail address: yhju@mail.ntust.edu.tw (Y.-H. Ju). 1385-8947/\$ - see front matter Ó 2013 Elsevier B.V. All rights reserved.
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07.048 substituent

11which enable them to protect human tissues from damages caused by oxygen or free radicals, and consequently reduce the risk of various diseases, and offer beneficial effect against cancer, cardiovascular disease, diabetes, and Alzheimer's disease

[2–7]. In starch extraction from sago pith, the sago pith slurry during sedimentation is browning due to the

30oxidation of phenolic compounds by the polyphenol oxidase (PPO) enzyme [8]. Phenolic compounds are located mainly in vacuoles while the PPO

is found in plastids. Browning is initiated with the disruption of cell due to

30cutting or bruising and the compartmentalization is lost. The

primary phenolic compounds related to browning are DL-epicatechin and D-catechin [8,9]. Pei-Lang et al. [9] reported that the content of phenolic compounds in sago pith depends on its growth stages and height; and the amount of phenolic compound extracted is less than 1%. Recovery of phenolic compounds from the waste effluents is beneficial, but it is a challenging process. Adsorption is efficient for removing

2organic matter from waste effluents. Activated carbons are the most widely used adsorbents due to their high adsorption capacities for organic pollutants

which are

2related to their high surface area, pore volume and porosity. The adsorption

capacities also strongly depend on the activation method and the nature of source materials.

2Activated carbon is an effective adsorbent for organic compounds removal, especially for phenolic

compounds [10–15].

40However, the utilization of activated carbon as adsorbent in the adsorption of

epicatechin and catechin has never been reported. The focus of this work was to evaluate the adsorption performance of activated carbon for the recovery of epicatechin and catechin in single and binary component systems. The surface area was obtained based on Brunauer, Emmet, and Teller (BET) measurements, while the pore volume and pore diameter of activated carbon were calculated from the adsorption experimental data using density functional theory (DFT). The effect of various variables, such as initial adsorbate concentrations, adsorption time and mass of adsorbent were studied using batch equilibrium technique.

27Pseudo first and second order kinetic models were used to correlate the adsorption dynamic or kinetic data. The

10adsorption equilibrium data was evaluated using Langmuir model

for single component, while

9modified extended-Langmuir model with incorporation of solute's selectivity factor

was used to study the correlation [16]. The feasibility and adsorption performance of activated carbon in recovering epicatechin and catechin was also tested in real effluent. 2. Experimental section

392.1. Chemicals HPLC grade (?)-epicatechin (purity

98 wt.%) and (+)-catechin hydrate (purity P

3998 wt.%) were obtained from Sigma Aldrich (Singapore) and was used without any further purification.

Pellet activated carbon (0.5–1.5) produced by Norit (The Netherlands) was used as the adsorbent. Double distilled water was used in adsorption experiments. 2.2. Characterizations of activated carbon Since

4 structural heterogeneity of activated carbon plays an important role during adsorption

therefore it is crucial to characterize the pore structure of activated carbon.

3 Pore structure of the activated carbon was characterized by nitrogen sorption technique in a Quadrasorb SI at

boiling point of nitrogen gas ($-196\text{ }^{\circ}\text{C}$). Prior to the nitrogen sorption measurement, the activated carbon was degassed in vacuum condition at $200\text{ }^{\circ}\text{C}$ for 48 h. The measurement of the nitrogen sorption isotherms (adsorption and desorption) were conducted

44 over a relative pressure (p/p_0) of 0.001–0.990. The BET surface area of the carbon was determined by

standard BET equation at a

28 relative pressure of 0.06–0.3. Total of pore volume of the activated carbon was

evaluated at the highest relative pressure (0.990).

4 At this relative pressure all pores

within the activated carbon were already completely filled with nitrogen gas.

4 Pore size distribution of the carbon was determined by standard DFT method with medium regularization.

2.3. Static adsorption experiments The adsorbate solution was prepared by mixing a known amount of adsorbate ((-)-epicatechin or (+)-catechin hydrate) with double distilled water to produce the desired initial

concentrations (100–150 mg/L). The adsorption was carried out isothermally in static mode at 303.15 K. A fixed amount of activated carbon was added to

16 a series of 250 mL glass-stoppered flasks each filled with 100 mL

diluted solutions. The glass stoppered flasks were then put

38 in a thermostatic shaker bath at 120 rpm for 1 h (the

equilibrium condition was achieved in about 40 min). The adsorption isotherm for binary systems of epicatechin and catechin was also obtained isothermally at 303.15 K, using the same procedure as for the single component adsorption. To study the adsorption kinetics, a series of 250 mL glass-stoppered flasks contained 100 mL of phenolic compound solution with certain initial concentration (100–250 mg/L) were prepared. Subsequently,

34 a known amount of activated carbon (0.2 g) was added into each glass-stopper flask. The

glass-stopper flasks were placed in thermostatic water-bath shaker for 150 min at 303.15 K. During kinetic experiments, at intervals of 10 min, one of glass-stopper flask was taken and the concentration of phenolic compound was analyzed. The adsorption of phenolic compounds from waste effluent was carried out using waste water from sago starch extraction. Briefly, sago pith was pulverized into coarse powder and soaked in water (1 h), and then filtered using Buchner funnel to obtain the waste effluent. One gram of activated carbon was added to 250 mL glass-stopper

16 flasks each filled with 100 mL

waste effluent. The glass stopper flask was placed

38 in a thermostatic shaker bath at 120 rpm for 1 h. The

initial and equilibrium concentrations of the single and binary solutions and waste effluent were analyzed

41 using a HPLC (Jasco HPLC PU-2089 plus) with a UV-vis detector (UV-2077 plus)

set at 280 nm. Separations were achieved in an Enduro C18, 5 μ m, 250 \times 4.6 mm. The

21flow rate was **0.8 ml/min**. The mobile phase used was

isocratic elution comprised of 3% acetic acid in double distilled water (60%) and acetonitrile (40%) for a total running time of 10 min. The contents of epicatechin and catechin were determined using calibration curves. The

14effect of pH on the adsorption of epicatechin and catechin was investigated in

single component systems. It was found that as the initial solution pH was reduced to pH 3, the solution concentration decreased up to 50% of the initial concentration. When the initial solution pH was increased to pH 10, the solution concentration also decreased up to 60–70% of the initial concentration. This is due to oxidation of the phenolic compounds. Therefore,

40the adsorption of epicatechin and catechin in single and binary component were

carried out at pH 7, which is similar to the waste effluent pH. 2.4. Calculations The amounts of epicatechin and catechin adsorbed onto activated carbon in single component system were calculated using the following equation: $q_e = \frac{C_0 - C_e}{m} V$ where C_0

25and C_e are the initial and equilibrium concentration of phenolic compounds in the liquid phase (mg/L), respectively, V is the volume of phenolic compounds solution (L), q_e is the amount of

metal ions adsorbed per unit of biosorbent mass (mg/g), and m is the mass of activated carbon

23(g). The equilibrium concentration of phenolic compound i

species ($C_{e,i}$) in binary mixture was calculated

23by the same manner using following mass balance equation: $q_e = \frac{C_0 - C_{e,i}}{m} V$

V where q_e

23, i is the equilibrium amount of solute i in the adsorbed phase

(mg/g), $C_{0,i}$ and C_e

3, i are the initial and equilibrium concentration **of** solute **i in the** liquid phase (mg/L), respectively. All data are **the** average results **of**

triplicates experiments. 408 F.E. Soetaredjo et al. / Chemical Engineering Journal 231 (2013) 406–413 3. Results and discussion 70 3.1. Activated carbon characterization 60 Analysis of nitrogen sorption isotherms and DFT-pore size distribution of the activated carbon used in this study indicates that the NORIT carbon possess both micropore and mesopore structures with micropore structure more dominant than mesopore as indicated by the rapid filling of pores at low relative pressure and followed by the gradual increase of adsorption at moderate relative pressure (S1). The existence of the mesopores

28 is indicated by the presence of hysteresis loop **between adsorption and desorption**

isotherms, an indication of capillary condensation in the mesopores. The

13 BET surface area, micropore volume, and total pore volume **of the**

activated carbon (NORIT nrs carbon Ea 0.5–1.5) were 890 m²/g, 0.431 cm³/g, and 0.506 cm³/g, respectively. The pore distribution of activated carbon represents a model of carbon internal structure which represents non-interacting of regularly shape model pores and

4 complex void spaces within the carbon. **The pore size distribution** of the carbon **is**

one of its most important characteristics of the carbon in studying adsorption equilibrium and kinetic. The

37 pore size distribution of the activated carbon confirms that the **carbon**

possesses both micropore and mesopore structure (S2). 50 40 30 q_e (mg.g⁻¹) 20 10 0 0 20 25 20 q_e (mg.g⁻¹) 15 10 (a) 40 60 C_e (mg.L⁻¹) 3.2. Adsorption isotherms 5 (b) The isotherm shape can be used as qualitative information

5 on the nature of the solute-surface interaction. In **the** case **of 0 adsorption of** phenolic compounds **from**

water, it is characterized 40 50 60 70 80 90 100 by initial region (high uptake at low concentration); and Langmuir C_e (mg.L⁻¹) model is best fitted for this case [17,18]. Based on the classification proposed by Giles et al. [17,18], the curve shape in Fig. 1 is class L2 Fig. 1. Adsorption isotherm of catechin and epicatechin onto activated carbon NORIT nrs carbon E_a 0.5–1.5 and Langmuir isotherm model (solid lines). according to the Giles classification

5 suggests that the aromatic ring adsorbed is parallel to the surface and no strong competition exists between the adsorbate and the solvent to occupy the adsorption

time consuming and much more difficult to obtain. Therefore it sites [12].

22 Langmuir isotherm is characterized by a plateau curve

will be convenient if the multi-component isotherm can be con- which indicates that equilibrium has been achieved. The Langmuir structured based on single adsorption isotherms. Various models isotherm was originally developed for describing gas adsorption have been developed for this purpose and one of the most widely phenomena but it has been extensively used for the correlation used is the extended Langmuir and its modified forms [16,22,23]. of

22 adsorption equilibrium data of various solute-sorbent interac- The

extended Langmuir model for a two-component system can tions in liquid phase [19–21]. The Langmuir isotherm model is gi- be expressed as: ven as follows: $q_e = \frac{1}{\frac{1}{q_m} + \frac{K_L C_e}{q_m}}$ where q_m

22 is the maximum adsorption capacity of the solid (mg /g) corresponding to monolayer surface coverage

and K_L is Langmuir $q_e = \frac{1}{\frac{1}{q_m} + \frac{K_L C_e}{q_m}}$ constants of adsorption affinity (L/mg). where $q_{m,i}$ and $C_{e,i}$ are the amount adsorbed and equilibrium con- Fig. 1 shows that the adsorption equilibrium data of phenolic concentration of component i , respectively. The parameters $q_{m,i}$ and compounds are well fitted by the Langmuir isotherm model and $K_{L,i}$ are the adsorption capacity of component i in the adsorbent the correlation coefficients for epicatechin and catechin are 0.97 and adsorption affinity of component i , respectively. All these and 0.94, respectively. The maximum adsorption capacity (q_m) of parameters are obtained from single adsorption isotherms. catechin (73.56 mg/g) obtained by fitting calculation using Sigma- In a multi-component system, competition for vacant adsorp- Plot 12.0 is higher than that of epicatechin (29.04 mg/g). The tion sites in the surface of adsorbent occurs not only between adsorption affinity parameter (K_L) of catechin and epicatechin ob- the same molecules but also between different adsorbate species. tained are 0.1257 L/mg and 0.041 L/mg, respectively. Higher This competition may act as the controlling factor for the adsorp- adsorption affinity parameter of catechin than epicatechin

indication process [16,23] and such phenomenon is not captured by original extended Langmuir model. Recently, Kurniawan et al. [23] the carbon. and Soetaredjo et al. [16] modified the extended Langmuir model Industrial Adsorption process usually involves multi-component by including fractional surface loading or coverage [23] and selection, therefore, information on adsorption equilibria in multi-component system is required for engineering design of the adsorption compounds, the fractional surface loading approach by Kurniawan process. Compare to the single component adsorption, reliable et al. [23] is more

10 suitable to represent the binary adsorption data experimental

multi-component adsorption isotherm data are more of this study. Since

37 competition between catechin and epicatechin molecules for vacant adsorption sites in the activated carbon occurred during the

adsorption process, catechin and epicatechin were adsorbed in certain amount, and the adsorption capacity of the activated for both adsorbates (binary) can be written as $q_{m,bin} = q_{m,1}h_1 + q_{m,2}h_2$ Here h_1 and h_2 are the fractional loading of catechin and epicatechin, respectively, and $h_1 + h_2 = 1$. The adsorption affinity parameter in Langmuir equation represents

27 how strong an adsorbate molecule is attracted onto the surface of the

adsorbent. In a binary system, the competition between different molecules weakens the adsorption potential

36 of the adsorbate toward the surface of adsorbent, therefore, the

affinity parameter of binary compounds can be written as: $K_{L,1(bin)} = \frac{1}{K_{L,1} + h_1}$ $K_{L,2(bin)} = \frac{1}{K_{L,2} + h_2}$ where $K_{L,1}$ and $K_{L,2}$ are the affinity parameter of catechin and epicatechin

12 in binary system, respectively. The final expressions of extended Langmuir model

with inclusion of fractional loading concept are $q_e = \frac{1}{\frac{1}{K_{L,1}q_{m,1} + h_1} + \frac{1}{K_{L,2}q_{m,2} + h_2}}$ $q_e = \frac{1}{\frac{1}{K_{L,1}q_{m,1} + h_1} + \frac{1}{K_{L,2}q_{m,2} + h_2}}$ The applicability of the modified extended Langmuir model was tested against the liquid phase binary adsorption data of catechin and epicatechin. The parameter δ_1 and δ_2 were obtained by the simultaneously fitting of Eqs. (9) and (10) using nonlinear least square method by minimizing the following objective function $\sum_{i=1}^n SSE_i = \sum_{i=1}^n (q_{exp,i} - q_{cal,i})^2$ where

q_{exp} and q_{cal} are the actual amount and the calculated values of catechin and epicatechin adsorbed by activated carbon, respectively; n is the total number of experimental data used in the fitting procedure. Fig. 2 shows the 3D plots of binary adsorption of catechin and epicatechin at various initial concentrations and the model calculated by Eqs. (9) and (10). The fitted parameters h_1 and h_2 with calculate values of q_m , $q_m(\text{bin})$, $KL_1(\text{bin})$, and $KL_2(\text{bin})$ are summarized in Table 1. In this work an improvement was made in the fitting procedure of modified extended Langmuir model by Kurniawan et al. [23]. Both Eqs. (9) and (10) were simultaneously fitted by nonlinear least square method with h_1 and h_2 as the fitted parameters, with boundary condition of the fitting procedure as $h_1 + h_2 = 1$. It is obvious that this fitting procedure

34 improved the applicability of the modified extended Langmuir model in correlating the binary experimental data of

catechin and epicatechin as indicated in Fig. 2. The maximum amount of adsorbates adsorbed in binary system ($q_m(\text{bin})$) was lower than in single system (q_m) due to the effect of molecular competition between both adsorbate molecules. This competition was accommodated in the modified extended Langmuir by taking into account of fractional loading coverage. The fractional coverage of catechin (h_1) and the maximum amount adsorbates adsorbed ($q_m(\text{bin})$) decreased with increasing initial epicatechin concentration as shown in Table 1. This result is consistent with the experimental data of catechin adsorption experimental data of epicatechin modified extended Langmuir model (b) g/g, 35 d b e o r s d 30 a d s u n 25 p o m o c 20 c i l n o h e 15 p 80 f o / L t g n u 10 o 40 m 140 a 130 120 Ce epicatechin 11, m0g/L 20 100 C e c 6 a t 0 e c h i n, m 90 adsorption experimental data of catechin adsorption experimental data of epicatechin modified extended Langmuir model Fig. 2. 3D plot of binary adsorption data and modified extended Langmuir model. (a)

24 Catechin 100 mg/L + epicatechin 100 mg/L, (b) catechin 100 mg/L + epicatechin 150 mg/L.

tent

3 with adsorption characteristics in single adsorption system that the adsorption capacity of epicatechin is less than

that of catechin, therefore increasing the concentration of epicatechin in binary system caused the interaction of adsorbate molecules to become even higher and reduce the value of $q_m(\text{bin})$. 3.3. Adsorption kinetics The effective design of an adsorption system requires information about the

1 rate at which the organic compounds removal takes place in a given sorbent /solution system.

Several adsorption kinetic

10 **models have been developed to describe the** adsorption **kinetics** in liquid phase. **The**

development of adsorption kinetic models mostly is based on fundamental approach to interfacial kinetics and its modifications [24]. Several models which have been developed 410 F.E. Soetaredjo et al. / Chemical Engineering Journal 231 (2013) 406–413 Table 1 Fitted and calculated parameters for catechin and epicatechin adsorption at several initial concentrations. Phenolic compounds mixture Fitted parameter Calculate parameter R² h₁ h₂ q_m,(bin), mg/g K_{L,1}(bin), L/mg K_{L,2}(bin), L/mg

24 **Catechin 100 mg/ L + Epicatechin 100 mg/L** Catechin **100 mg/ L** + Epicatechin **150 mg/L**

0.632 0.368 0.524 0.476 57.1 0.0699 52.2 0.0499 0.0075 0.0139 0.998 0.997 based on this approach are Langmuir kinetics, first and second order, reversible first and second order, and irreversible reactions [24]. Another approach which is used to develop the kinetic models is based on assuming that surface reaction is the rate controlling step in sorption. Many simple and compact models have been proposed based on this approach and the famous and

43 **widely used models** are **pseudo-first** (Lagergren) **and pseudo-second order** [24]. **Pseudo-first order model** which is also known as

Lagergren equation is the earliest empirical model describing the rate of adsorption in a liquid phase system [25]. The differential form of this model is $\frac{dq}{dt} = k_1(q_e - q)$ The

36 **integration of Eq. (12) with initial** conditions: **$q(t) = 0$ at $t = 0$**

gives $q = q_e(1 - \exp(-k_1 t))$ where

21 **$q(t)$ is the amount of** adsorbate **adsorbed at time t (mg /g), and k_1 is the rate constant (1/min).**

Pseudo-second order was first proposed by Blanchard et al. [26]. This model assumes that

1 **direct adsorption/desorption (seen as chemical reaction) is the**

rate controlling step in the overall sorption kinetics [24], and has the differential form as $\frac{dq}{dt} = k_2(q_e - q)^2$. Here

k_2 is rate constant of pseudo-second order (g/mg min). Integration of Eq. (14) gives **the**

following result $\frac{1}{q_e - q} \ln \frac{q_e(q_e - q_0)}{q(q_e - q_0)}$. To obtain the parameters of pseudo-first and pseudo-second order equations, non-linear least square method was employed by fitting the kinetic experimental data with Eqs. (13) and (15)

and the results are summarized in Table 2. **Figs. 3 and 4**

present the plots of catechin and epicatechin uptake as the function of time, respectively. The

symbols represent the kinetic experimental **data while the solid lines represent**

the pseudo-first and pseudo-second order kinetic models. As shown in Table 2, the values of R^2 obtained from the fitting of kinetic experimental data using both models were similar, however the pseudo-second order kinetic model represents kinetic experimental data better than the pseudo-first order since the predicted amount of adsorbate adsorbed in equilibrium condition was closer to that of the measured one. This is confirmed by the fact that from the values of fitting parameter q_e of pseudo-second order model agree with the values of q_e from kinetic experimental data. As mentioned by Plazinski et al. [24] that one of the potential advantages of using pseudo-second order kinetic model to correlate the adsorption kinetic experimental data

is its small sensitivity to the influence of the random experimental error,

especially for the fitting parameter q_e . Another advantage of using the pseudo-second order is that the value of fitted parameter q_e can be obtained from the independent plot of kinetic adsorption experimental data [24,27]. In many adsorption kinetic studies [24,27–29],

failure of the pseudo-first order in correlating adsorption **kinetic experimental data is due to** the **theoretical**

interpretations of this equation. In pseudo-first order it is based on the common

assumption that the overall sorption process is controlled by the rate of adsorption **/desorption processes** which is **seen** as **a chemical reaction on the**

surface of

adsorbent [24]. However in many cases, the rate of diffusion of solute into interior of the adsorbent also controls the adsorption mechanism and this phenomenon is not captured by the pseudo-first order model. Table 2, Figs. 3 and 4 clearly indicate that the parameter k_2

1 plays a role as time scaling factor.

At high k_2 , shorter

1 time is required to reach equilibrium.

As can be seen in Table 2, k_2 decreased with increasing initial concentration. A simple correlation between of k_2 and initial concentration of the adsorbate (C_0) is given as follows: $k_2 = k_{2,0} \exp(-bC_0)$ where $k_{2,0}$, a and b are empirical constants. Plots of C_0 versus k_2 are given in Fig. 5. The fitted empirical constants for catechin are $k_{2,0} = 0.0013$ g/mg min, $a = 0.0194$ g/mg min, and $b = 0.0092$ L/mg, while for epicatechin are $k_{2,0} = 0.0004$ g/mg min, $a = 0.0147$ g/mg min, and $b = 0.0207$ L/mg. Table 2 Fitted parameters of pseudo-first and pseudo-second order of catechin and epicatechin adsorption onto activated carbon NORIT nrs carbon E_a 0.5–1.5. Phenolic compound C_0 , mg/L

26 q_e , mg/g, experiment Pseudo-first order Pseudo-second order k_1 , 1/min q_e , mg/g R^2 k_2 , g/mg min q_e , mg/g R^2

Catechin 101.3 122.5 139.8 158.7 203.6 251.9 Epicatechin 100.5 118.4 141.2 161.5 201.7 252.3 43.3 43.9
 44.5 44.7 45.3 46.1 0.0631 0.0471 0.0441 0.0337 0.0275 0.0185 21.2 21.9 23.2 25.4 28.2 28.9 0.1037
 0.1096 0.0974 0.0795 0.0708 0.0656 39.1 38.6 38.3 38.2 35.6 33.6 0.996 0.997 0.998 0.998 0.996 0.994
 19.6 22.2 21.1 22.5 25.4 26.5 0.993 0.996 0.996 0.995 0.994 0.995 0.0022 0.0014 0.0013 0.0008 0.0007
 0.0004 43.4 44.3 44.3 46.1 44.2 45.1 0.0086 0.0083 0.0069 0.0053 0.0040 0.0034 20.9 23.6 22.8 24.5 27.9
 29.3 0.997 0.998 0.999 0.998 0.997 0.996 0.994 0.998 0.996 0.996 0.997 0.995 50 (a) 40 qt (mg.g-1) 30 20
 10 0 0 20 40 60 80 100 120 140 160 T (min) 50 (b) 40 qt (mg.g-1) 30 20 10 0 0 20 40 60 80 100 120 140
 160 T (min) Fig. 3. Kinetic adsorption experimental data of catechin onto activated carbon NORIT nrs carbon E_a 0.5–1.5 at various initial concentration

7 and plots of (a) pseudo- first order, and (b) pseudo-second order.

3.4. Recover of phenolic compounds in sago waste effluent To verify the potential application of recovering phenolic compounds from sago waste effluent by adsorption, adsorption experiments using real sago waste effluent were also conducted. The adsorption of catechin and epicatechin from real sago waste effluent was conducted isothermally at 303.15 K under static condition. Prior to the adsorption experiment, the sago waste effluent was filtered to remove solid particles. The initial concentration of catechin and

epicatechin in sago waste effluent was 321.8 mg/L and 158.2 mg/L, respectively. The recovery of catechin and epicatechin in sago waste effluent

13 is given in Table 3. From Table 3 it can be seen that the

recovery of both phenolic compounds increases with increasing activated carbon mass due to the increase of adsorption active sites. At an adsorbent dose of 2%, the

19 recovery of catechin and epicatechin from sago waste effluent

was 85% and 53%, respectively. It can be seen that the amount of phenolic compounds adsorbed by the activated carbon from real waste sago effluent (Table 3) is less than that of the synthetic one (Fig. 2). In waste sago effluent, in addition to catechin and epicatechin other compounds such as protein and soluble starch are also present which compete for adsorption active sites

29 resulting in the observed decrease of adsorption capacity

toward catechin and epicatechin.

29 Reusability of adsorbent is crucial in industrial application. The

recovery of catechin and epicatechin from activated carbon was performed by supercritical CO₂ at 313.15 K and 26 MPa under static and dynamic conditions. Details about the supercritical system can be found elsewhere [30]. At static condition, the recovery of catechin from activated carbon was only 1.2% while for epicatechin it was 14.5%. This low recovery is consistent with the solubility

15 (mg.g⁻¹) 15 10 5 0 0 20 40 60 80 100 120 140 160 T (min)

30 (b) 25 20 qt

15 (mg.g⁻¹) 15 10 5 0 0 20 40 60 80 100 120 140 160 T (min) Fig. 4.

Kinetic adsorption experimental data of epicatechin onto activated carbon NORIT nrs carbon Ea 0.5–1.5 at various initial concentration

7 and plots of (a) pseudo- first order, and (b) pseudo-second order. data

of both phenolic compounds in supercritical CO₂ under static condition [30,31]. In order to increase the recovery of catechin and epicatechin in supercritical CO₂, dynamic extraction experiments were also conducted with flow rates of CO₂ from 0.1 to 1 kg/h. By increasing CO₂ flow rate from 0 kg/h (static mode) to 0.1 kg/h, the recovery increased from 1.2% to 2.3% and from 14.5% to 27% for catechin and epicatechin, respectively. Maximum recovery (catechin 9.7% and epicatechin 87.3%) was achieved

42at a CO₂ flow rate of 0.7 kg/h.

Further increase of CO₂ flow rate to 1 kg/h had negligible effect on the recovery both compounds. 0.010 0.008 Catechin Epicatechin k₂, g/mg.min 0.006 0.004 0.002 0.000 80 100 120 140 160 180 200 220 240 260 Initial concentration, mg/L Fig. 5. Correlation between Co and k₂. 412 F.E. Soetaredjo et al. / Chemical Engineering Journal 231 (2013) 406–413 Table 3 Adsorption of catechin and epicatechin from sago waste effluent. Co,1 catechin, mg/ Co,2 epicatechin, mg/ m, mass of activated carbon, L L g Ce,1, mg/ Ce,2, mg/ L L qe,1, mg/ qe,2, mg/ % Recovery g g catechin % Recovery epicatechin 321.8 158.2 0.112 0.483 0.689 0.921 1.104 1.328 1.573 1.921 274.4 143.3 141.2 113.8 110.3 103.8 99.8 100.2 95.5 96.4 85.4 93.1 71.2 87.4 48.3 74.2 42.3 13.3 37.4 9.2 30.7 7.9 24.1 6.3 20.5 5.6 17.8 4.9 15.9 4.5 14.2 4.3 14.7 9.4 56.1 28.1 65.7 34.4 68.9 36.7 70.3 39.1 73.5 41.1 77.7 44.7 84.8 52.3 Note: volume of sago waste effluent 100 mL. Low recovery of phenolic compound, especially catechin is caused by the difference in the polarity between the solute (phenolic compound) and the solvent (supercritical CO₂). Catechin is highly polar while supercritical CO₂ is non-polar. In order to increase the polarity of supercritical CO₂, ethanol as co-solvent (5–25%) was used in this study. By adding 5% ethanol

42at a CO₂ flow rate of 0.7 kg/h,

the recovery of catechin increased almost two fold (from 9.7% to 18.2%), and while for epicatechin the recovery reached 95.7%. Increasing the percentage of co-solvent increased the recovery of both compounds. For epicatechin the maximum recovery was 99.2% at 20% of co-solvent. Further increase in co-solvent concentration had negligible effect on the recovery of epicatechin. The un-recovered epicatechin may be strongly bound on carbon surface or epicatechin molecule may be adsorbed inside small or ultra micropore so it could not be desorbed by supercritical CO₂. At 30% co-solvent, the recovery of catechin was 93.1%. Since further increase of co-solvent concentration changed supercritical condition into subcritical, therefore maceration (303.15 K, 24 h) was used to obtain maximum removal of catechin (97.8%) and epicatechin (99.4%). 4. Conclusion The

19recovery of catechin and epicatechin from sago waste

effluent by adsorption onto activated carbon followed

9by supercritical CO₂ desorption was demonstrated in this work. To study the

kinetic and equilibria of adsorption, synthetic aqueous catechin and epicatechin solution was used instead of sago waste effluent. The adsorption equilibria and kinetic were obtained isothermally at 303.15 K in static mode condition. The adsorption equilibria of single and binary system were modeled using Langmuir and modified extended Langmuir, respectively. It was found that both of

29 **models could represent the experimental data well. In** adsorption kinetic experiments, **the**

well-known

9 **pseudo-first and pseudo-second models were** employed **to correlate the experimental data. The pseudo-second**

model gave better representation of experimental data than the pseudo-first model. The effect of initial concentration on the value of fitting parameters k_1 and k_2 was also studied. The results indicate that k_2

1 **plays a role as time scaling factor,**

at high k_2 , shorter

1 **time is required to reach equilibrium.**

Adsorption using real sago waste effluent followed by supercritical CO₂ extraction indicates that this method has potential application in recovery catechin and epicatechin in industrial scale. Ethanol as co-solvent was required to obtain high recovery of catechin from the activated carbon. References [1] M.A. Soobrattee, V.S. Neerghen, A. Luximon-Ramma, O.I. Aruoma, T. Bahorun, Phenolics as potential antioxidant therapeutic agents: mechanism and actions, *Mutat. Res.* 579 (2005) 200–213. [2] L. Yu, K. Zhou, Antioxidant properties of bran extracts from “platte” wheat grown at different locations, *Food Chem.* 90 (2004) 311–316. [3] P. Vitaglione, A. Napolitano, V. Fogliano, Cereal dietary fibre: a natural functional ingredient to deliver phenolic compounds into the gut, *Trends Food Sci. Technol.* 19 (2008) 451–463. [4] A.R. Rechner, M.A. Smith, G. Kuhnle, G.R. Gibson, E.S. Debnam, S.K.S. Srari, K.P. Moore, C.A. Rice-Evans, Colonic metabolism of dietary polyphenols: influence of structure on microbial fermentation products, *Free Radical Biol. Med.* 36 (2004) 212–225. [5] A.R. Rechner, G. Kuhnle, P. Bremner, G.P. Hubbard, K.P. Moore, C.A. Rice-Evans, The metabolic fate of dietary polyphenols in humans, *Free Radical Biol. Med.* 33 (2002) 220–235. [6] O. Pourali, F.S. Asghari, H. Yoshida, Production of phenolic compounds from rice bran biomass under subcritical water conditions, *Chem. Eng. J.* 160 (2010) 259–266. [7] H. Epstein, Cosmeceuticals and polyphenols, *Clin. Dermatol.* 27 (2009) 475–478. [8] S.M. Anthonysamy, N.B. Saari, K. Muhammad, F.A. Bakar, Browning of sago (*Metroxylon sagu*) pith slurry influenced by holding time, pH and temperature, *J. Food Biochem.* 28 (2004) 91–99. [9] A.T. Pei-Lang, A.M.D. Mohamed, A.A. Karim, Sago starch and composition of associated components in palms of different growth stages, *Carbohydr. Polym.* 63 (2006) 283–286. [10] M. Ahmaruzzaman, Adsorption of phenolic compounds on low-cost adsorbents: a review, *Adv.*

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